5,6-UNSATURATED HEXOFURANOSYL GLYCOSIDES AND 5',6'-UNSATURATED HEXOFURANOSYL NUCLEOSIDES

LEON M. LERNER

Department of Biochemistry, State University of New York, Downstate Medical Center, Brooklyn, New York 11203 (U. S. A.)

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ABSTRACT

Methyl 5,6-dideoxy-2,3-O-isopropylidene- α -D-lyxo-hex-5-enofuranoside, prepared from methyl 2,3-O-isopropylidene-5,6-di-O-methylsulfonyl- α -D-mannofuranoside with sodium iodide in 2-butanone, was acetolyzed and the product coupled with 6-benzamidochloromercuripurine by the titanium tetrachloride method. Removal of the N-benzoyl group with picric acid afforded 9-(2,3-di-O-acetyl-5,6-dideoxy- β -D-xylo-hex-5-enofuranosyl)adenine. In a similar manner, methyl 5,6-dideoxy-2,3-O-isopropylidene- α -L-lyxo-hex-5-enofuranoside was prepared from L-mannose and converted into 9-(2,3-di-O-acetyl-5,6-dideoxy- β -L-xylo-hex-5-enofuranosyl)adenine, further de-esterified to give the free nucleoside. 2,3:5,6-Di-O-isopropylidene- α -L-mannofuranosyl chloride, prepared from L-mannose, gave 9-(2,3-O-isopropylidene- α -L-mannofuranosyl)adenine, hydrolyzed into 9- α -L-mannofuranosyladenine. Treatment with methanesulfonyl chloride gave the 5',6'-dimethanesulfonate, which gave with sodium iodide in acetone the 5',6'-unsaturated nucleoside, further hydrolyzed into 9-(5,6-dideoxy- α -L-lyxo-hex-5-enofuranosyl)adenine.

INTRODUCTION

Unsaturated pentofuranosyl nucleosides have been prepared from purine and pyrimidine nucleosides having the double-bond located between the 1',2' (Ref. 1), 2',3' (Ref. 2), and 3',4' (Ref. 3) positions. All of these aldofuranosyl nucleosides have the site of unsaturation within the carbohydrate ring. Nucleosides having an exocyclic 4',5' double-bond have been prepared^{4,5} and have also been isolated as elimination products during attempts^{6,7} at nucleophilic substitution at C-5'. One of these nucleosides, 9-(5-deoxy- β -D-erythro-pent-4-enofuranosyl)adenine (1), exhibited an anti-bacterial activity⁴ similar to that of the ketohexofuranosyl nucleoside antibiotic decoyinine (Angustmycin A, 2), a compound also reported to have antitumor activity⁸. It was of interest, therefore, to explore further the significance of exocyclic double-bonds of nucleosides in relation to the biological, especially antitumor and antibacterial activity.

$$CH_{2} \xrightarrow{\text{PO}} Ade$$

$$1 \text{ R} = H$$

$$2 \text{ R} = CH_{2}OH$$

$$3$$

$$CH_{2} = CH_{2}OH$$

$$4$$

$$Ade = Adenine$$

$$3$$

$$3$$

$$CH_{2} = CH_{2}OH$$

$$4$$

$$5 \text{ R} = Ac$$

$$6 \text{ R} = H$$

In past work from this laboratory^{9,10}, hexofuranosyl nucleosides were synthesized that are unsaturated between C-5' and -6' and, therefore, the site of the double bond was located outside the furanose ring. One of these compounds, 9-(5,6-dideoxy-α-D-lyxo-hex-5-enofuranosyl)adenine (3) had antibacterial activity and another related nucleoside derivative, 9-(2,3-di-O-acetyl-5,6-dideoxy- β -D-xylo-hex-5-enofuranosyl)adenine (4) was active against leukemia L1210 cells in vitro11. It was desirable to improve the synthesis of 4 and to synthesize other related hexofuranosyl nucleosides. The previous¹⁰ two syntheses of 4, both starting from 1,2:5,6di-O-isopropylidene-α-D-glucofuranose were time-consuming and tedious. Furthermore, the method of olefin formation from the preformed nucleoside, although smoothly performed in the preparation⁹ of 3, was not achieved with a good yield in the case of 4, because of competing anhydronucleoside formation during the sodium iodide-acetone reaction at elevated temperatures. Indeed, the synthesis of 4 was best achieved from a preformed, unsaturated carbohydrate derivative¹⁰. This report describes the preparation of 5,6-unsaturated glycosides and, from them, the desired unsaturated nucleosides.

RESULTS AND DISCUSSION

The preparation of the enantiomeric 9-(2,3-di-O-acetyl-5,6-dideoxy- β -xylohex-5-enofuranosyl)adenines (4 and 5) started from D- and L-mannose, rather than from D- and L-glucose. Recently, a remarkably simple and rapid preparation of methyl 2,3-O-isopropylidene-5,6-di-O-methylsulfonyl- α -D-mannofuranoside (7) from D-mannose was reported¹², and this preparation has been scaled up many times and utilized in the synthesis of 6-deoxyhexoses¹³. In a manner similar to that used with D-mannose, L-mannose was converted^{12,13} into methyl 2,3-O-isopropylidene-5,6-di-O-methylsulfonyl- α -L-mannofuranoside (10). Treatment of 7 and 10 with sodium iodide in boiling 2-butanone gave the 5,6-unsaturated glycosides, methyl 5,6-dideoxy-2,3-O-

isopropylidene-α-D(and L)-lyxo-hex-5-enofuranoside (8 and 11), respectively. This reaction required nearly 24 h for completion, and the products 8 and 11 were obtained in very high yields after purification by distillation. Although 8 does not appear to have been described in the literature, 11 has previously been prepared from methyl 2,3-O-isopropylidene-5-O-p-tolylsulfonyl-α-L-rhamnofuranoside as a minor product during attempts at nucleophilic substitution¹⁴.

For the conversion of the glycosides 8 and 11 into the enantiomeric 1,2,3-tri-O-acetyl-5,6-dideoxy-xylo-hex-5-enofuranoses (9 and 12), the conditions, first described by Jerkeman¹⁵, have recently been developed into a workable synthetic procedure^{13,16}. The limitations of this epimerization have been studied^{17–19} and at least two mechanisms^{15,17} have been proposed to explain it. The triacetates 9 and 12 were utilized as crude syrups in the synthesis of the nucleoside 4 and its L form 5.

The number of coupling procedures which can be utilized for the preparation of nucleosides are obviously limited by the presence of the unsaturated group and for this reason the titanium tetrachloride procedure²⁰ was used, although presumably the fusion method ²¹ may be an alternative choice. Coupling of syrupy 9 and 12 with 6-benzamidochloromercuripurine in the presence of titanium tetrachloride in 1,2-dichloroethane gave crude products, isolated as picrates²², which were converted²³ into the crystalline nucleosides 4 and 5. Removal of the acetyl groups of 5 in methanolic sodium methoxide gave 9-(5,6-dideoxy- β -L-xylo-hex-5-enofuranosyl) adenine (6) which, like its enantiomer¹⁰, was obtained only as an amorphous powder.

An attempt to prepare nucleoside 3 from 8 failed primarily because conditions could not be found that would selectively remove the isopropylidene group and afford the methyl glycoside. The original intention was to block OH-2 and -3 as the benzoic esters and proceed to form the nucleoside in a manner similar to that recently used to prepare 9-\alpha-D-rhamnofuranosyladenine²⁴. Application of this route to form 9-(5,6-dideoxy-\alpha-L-lyxo-hex-5-enofuranosyl)adenine (19) was attempted in order to reduce the number of derivatives of L-mannose that had to be prepared. However, since it was not successful, the synthesis of 19 was performed as previously described^{9,25} for 3. L-Mannose was converted in two steps into 2,3:5,6-di-O-isopropylidene-\alpha-L-mannofuranosyl chloride (14), which was purified by distillation. The coupling reaction was performed by the method of Davoll and Lowy²⁶ and, after

selective removal of the N-benzoyl and 5',6'-isopropylidene groups, 9-(2,3-O-isopropylidene- α -L-mannofuranosyl)adenine (15) was obtained in crystalline form. Mesylation of 15 gave syrupy 17 which was treated with sodium iodide in acetone to afford 9-(5,6-dideoxy-2,3-O-isopropylidene- α -L-lyxo-hex-5-enofuranosyl)adenine (18). Acid hydrolysis of 18 gave 19, which had been previously isolated as a minor elimination product from 9-(2,3-O-isopropylidene-5-O-p-tolylsulfonyl- α -L-rhamnofuranosyl)adenine⁷. In addition, acid hydrolysis of 15 gave 9- α -L-mannofuranosyladenine (16). Weak antitumor activity^{11,27} and inhibition of adenine phosphoribosyl transferase of Ehrlich ascites tumor cells²⁸ has been observed with the D enantiomer. An alternative and less expensive route to 18 may proceed from D-gulose via the known 9-(2,3-O-isopropylidene- β -D-gulofuranosyl)adenine²⁹, if future biological studies should warrant large-scale synthesis.

EXPERIMENTAL

General methods. — Melting points were determined with a Kofler hot-stage, and optical rotations with a Rudolph polarimeter. I.r. spectra were recorded with a Perkin-Elmer Model 21 spectrophotometer and n.m.r. spectra with a Varian T-60A spectrometer. Moist organic solutions were dried with anhydrous magnesium sulfate, and evaporations were performed in vacuo in a rotary evaporator at a bath temperature of 40-45° unless otherwise stated. Elementary analyses were performed by the Spang Microanalytical Laboratory, Ann Arbor, Michigan.

L-Mannose. — Most of the L-mannose utilized in this study was obtained by reduction of L-mannono-1,4-lactone (Pfanstiehl Labs. Inc., Waukegan, Ill. 60085) with sodium borohydride under acidic conditions following the general methodology³⁰ and slowly crystallized from 2-propanol-methanol mixtures. In some cases, as in the preparation of 10, the syrupy form was satisfactory for use. Small amounts of L-mannose were also obtained from Pfanstiehl Labs.

Methyl 5,6-dideoxy-2,3-O-isopropylidene- α -D-lyxo-hex-5-enofuranoside (8). — Methyl 2,3-O-isopropylidene-5,6-di-O-methylsulfonyl- α -D-mannofuranoside¹² (7,

25 g) was treated with sodium iodide (70 g) in 2-butanone (550 ml) for 24 h at reflux. The solvent was evaporated (35°) and the residue was partitioned between chloroform (200 ml) and a sodium thiosulfate solution (250 ml, 10%). The chloroform layer was washed once more with a sodium thiosulfate solution (250 ml) and then with water (250 ml), dried, and evaporated. The residual oil was distilled to yield 12.4 g (96%), b.p. 87–95° (3 mm Hg), $[\alpha]_D^{22} + 27.8^\circ$ (c 2.76, chloroform); the i.r. and n.m.r. spectra were identical to those of the L form 11.

Anal. Calc. for C₁₀H₁₆O₄: C, 59.98; H, 8.06. Found: C, 60.13; H, 8.13.

 $9-(2,3-Di-O-acetyl-5,6-dideoxy-\beta-D-xylo-hex-5-enofuranosyl)$ adenine (4). — Methyl 5,6-dideoxy-2,3-O-isopropylidene- α -D-iyxo-hex-5-enofuranoside (8, 1.0 g) was dissolved in a mixture of glacial acetic acid (30 ml) and acetic anhydride (3 ml), chilled in an ice-bath, and sulfuric acid (1.65 ml) was added dropwise. The mixture was kept at room temperature for 65 h and then poured into ice-water (125 ml) and stirred. When the ice had melted, the product was extracted with chloroform (3×25 ml) and the chloroform solution was washed with water (100 ml), saturated sodium hydrogencarbonate (100 ml), and again with water. The chloroform solution was dried and evaporated. Several additions and evaporations of benzene gave the triacetate 9 as a clear, colorless oil (1.03 g, 76%); the n.m.r. spectrum showed that the methoxyl and isopropylidene groups had been removed and replaced by acetyl groups (methyl peaks centered at δ 2.08).

The oil (9) was dissolved in 1,2-dichloroethane (100 ml) and added to a mixture containing 6-benzamidochloromercuripurine (2.23 g) and Celite-545 (2.2 g), and 20 ml of the solvent was removed by distillation. Titanium tetrachloride (0.35 ml), dissolved in 1,2-dichloroethane (20 ml), was added and the mixture was heated at reflux for 22 h. The reaction mixture was cooled to room temperature and treated with saturated sodium hydrogencarbonate (100 ml). After 1 h of stirring, the mixture was filtered and the filter cake was washed with hot 1,2-dichloroethane (70 ml). The organic layer was separated, evaporated to dryness, and the residue was dissolved in chloroform (70 ml). This was washed with 30% aqueous potassium iodide (2 × 50 ml) and water (100 ml), dried, and evaporated to a syrup, which was dissolved in ethanol (10 ml), treated with 10% ethanolic picric acid (20 ml), and heated at reflux²². After 5 min, yellow crystals began to form, and boiling was continued for another 5 min. The flask was kept at room temperature for several h. The picrate was filterd off, dissolved in 80% aqueous acetone (150 ml), and treated with Bio-Rad AG1-X8 (CO_3^{2-}) ion-exchange resin to remove the picrate ion²³. The mixture was also treated with Darco G-60 charcoal, filtered, and the solution evaporated to afford a white foam which was treated by several additions and evaporation of ethanol until crystallization occurred. Product 4 was crystallized from ethanol in two crops to give 254 mg (20%), m.p. 200-201° with prior softening beginning at \sim 196°; the i.r. spectrum was identical to that of the previous preparation¹⁰, m.p. 201-203°

Methyl 2,3-O-isopropylidene-5,6-di-O-methylsulfonyl- α -L-mannofuranoside (10). — The preparation of 10 was virtually the same as that described 12,13 for the D form (7). From syrupy L-mannose (26.6 g) was obtained 22.5 g (39%) of large, white

needles, crystallized from methanol, m.p. 147–148.5°, $[\alpha]_D^{25}$ – 32.6° (c 1.80, chloroform). When 7 was crystallized from methanol, it had m.p. 147–148°, $[\alpha]_D^{25}$ + 33.5° (c 1.39, chloroform)¹³. In addition, the i.r. and n.m.r. spectra of 10 and 7 were identical.

Anal. Calc. for $C_{12}H_{22}O_{10}S_2$: C, 36.91; H, 5.68; S, 16.43. Found: C, 36.93; H, 5.69; S, 16.38.

Methyl 5,6-dideoxy-2,3-O-isopropylidene- α -L-lyxo-hex-5-enofuranoside (11). — Treatment of 10 with sodium iodide in 2-butanone, as described for 8, yielded 9.41 g (82%) of an oil after distillation, b.p. 53-54° (0.8 mm Hg), $[\alpha]_D^{25} - 27.6$ ° (c 2.69, chloroform); lit. b.p. 76° (bath, 3.2 mm Hg), $[\alpha]_D^{20} - 23.5$ ° (c 1.4, chloroform) and b.p. 40-42° (bath, 12 mm Hg), $[\alpha]_D - 28 \pm 2$ ° (c 0.7, chloroform); the n.m.r. and i.r. spectra also had the peaks previously reported.

Anal. Calc. for C₁₀H₁₆O₄: C, 59.98; H, 8.06. Found: C, 59.95; H, 8.16.

9-(2,3-Di-O-acetyl-5,6-dideoxy- β -L-xylo-hex-5-enofuranosyl)adenine (5). — Compound 11 (4 g) was acetolyzed as described for the D form 8. A clear, colorless syrup 12 (3.82 g, 70%) was obtained. It was coupled in a mixture containing 6-benzamidochloromercuripurine (8.3 g), Celite-545 (8.3 g), titanium tetrachloride (1.3 ml), and 1,2-dichloroethane (400 ml), as described for the preparation of 4. Product 5 was isolated via the picrate to afford 748 mg (15%). An analytical sample was recrystallized from methanol, m.p. 199-201° with softening starting at about 194°, $[\alpha]_D^{23}$ -21.9° (c 1.30, chloroform); the i.r. and n.m.r. spectra were identical to those of 4, which was previously reported¹⁰ to have $[\alpha]_D^{23} + 22.5^{\circ}$ (c 1.12, chloroform).

Anal. Calc. for $C_{15}H_{17}N_5O_5$: C, 51.87; H, 4.93; N, 20.16. Found: C, 51.99; H, 4.93; N, 20.17.

9-(5,6-Dideoxy- β -L-xylo-hex-5-enofuranosyl)adenine (6). — The blocked nucleoside 5 (146 mg) was treated with 0.1m methanolic sodium methoxide (25 ml) for 3 h at room temperature. Neutralization was effected with CG-120 (H⁺) ion-exchange resin, which was removed by filtration, and the methanol evaporated. The syrupy residue was dissolved in acetone (20 ml), filtered, and evaporated. The residue was repeatedly dissolved in acetone and evaporated until a hard, white foam (74 mg) was obtained, which was pulverized and dried under high vacuum; the i.r. spectrum was identical to that of the D enantiomer¹⁰.

Anal. Calc. for $C_{11}H_{13}N_5O_3$: C, 50.18; H, 4.98; N, 26.60. Found: C, 48.95; H, 4.96; N, 26.07.

A picrate was prepared in water, m.p. 204–207° (dec.). The p form¹⁰, prepared in methanol and recrystallized, had m.p. 209–211° (dec.).

2,3:5,6-Di-O-isopropylidene-L-mannofuranose (13). — To a solution of dry acetone (150 ml) and sulfuric acid (3.5 ml) was added L-mannose (5 g). The reaction mixture was stirred at room temperature and a clear solution was obtained within about 1 h. After a total reaction time of 3.5 h, the slightly yellow solution was chilled in an ice-bath and treated with conc. ammonium hydroxide (13.5 ml). The precipitate was removed by filtration on a pad of Celite and the filtrate was evaporated to dryness. A solid residue formed which was dissolved in chloroform, dried, and evaporated to a

small volume. Petroleum ether (b.p. $60-110^{\circ}$) was added to incipient turbidity and crystallization occurred, affording 5.67 g, A second crop gave 0.53 g for a total yield of 86%; m.p. $121.5-122.5^{\circ}$, $[\alpha]_{\rm D}^{24}-15.7^{\circ}$ (c 2.5, ethanol); the i.r. spectrum was identical to that of the D form²⁵, m.p. 122° , $[\alpha]_{\rm D}^{19}+16.6^{\circ}$ (c 2.5, ethanol). The L isomer 13 was originally synthesized by Iwadare³¹, m.p. $122-122.5^{\circ}$, $[\alpha]_{\rm D}-27^{\circ}$ (acetone).

Anal. Calc. for C₁₂H₂₀O₆: C, 55.37; H, 7.74. Found: C, 54.58; H, 7.45.

2,3:5,6-Di-O-isopropylidene-L-mannofuranosyl chloride (14). — This compound was prepared from 13 (19 g), as described for the D enantiomer²⁵, with thionyl chloride in pyridine³². The oil was distilled to give 11.6 g (57%), b.p. 135-137° (2.4 mm Hg).

Anal. Calc. for $C_{12}H_{19}ClO_5$: C, 51.70; H, 6.09; Cl, 12.72. Found: C, 51.76; H, 6.30; Cl, 12.65.

9-(2,3-O-Isopropylidene- α -L-mannofuranosyl)adenine (15). — The chloride 14 (11.5 g) was treated with 6-benzamidochloromercuripurine (19.8 g) in xylene as described for the D enantiomer²⁵. After selective removal of the N-benzoyl and 5,6-O-isopropylidene groups, product 15 (6.41 g) was crystallized from water. An additional crop (0.72 g) was obtained from the mother liquor via the picrate for a total yield of 52%; m.p. 247-250° with softening at 240°, $[\alpha]_D^{24}$ - 34° (c 1.16, 0.1M hydrochloric acid); lit. (D form)²⁵: m.p. 249-250°, $[\alpha]_D^{21}$ + 32.5° (c 1.26, 0.1M hydrochloric acid); the i.r. spectrum of 15 was identical to that of its enantiomer.

Anal. Calc. for $C_{14}H_{19}N_5O_5$: C, 49.85; H, 5.68; N, 20.76. Found: C, 49.69; H, 5.73; N, 20.62.

9- α -L-Mannofuranosyladenine (16). — The free nucleoside was obtained by hydrolysis of 15 (1.5 g) in hot, 25% aqueous acetic acid to yield 843 mg (64%), m.p 239-241°, $[\alpha]_D^{24}$ -74.2° (c 1.32, M hydrochloric acid); the D form²⁵ had m.p. 237-237.5°, $[\alpha]_D^{21}$ +74.8° (c 3.05, M hydrochloric acid); the i.r. spectrum of 16 was identical to that of the D form.

Anal. Calc. for $C_{11}H_{15}N_5O_5$: C, 44.45; H, 5.09; N, 23.56. Found: C, 44.36; H, 5.12; N, 23.33.

9-(5,6-Dideoxy-2,3-O-isopropylidene-α-L-lyxo-hex-5-enofuranosyl)adenine (18). — Treatment of 15 (2.28 g) with methanesulfonyl chloride in pyridine, as described for the D enantiomer⁹, gave 17 as a tan foam. Elimination was effected with sodium iodide in acetone at 100° to give 562 mg (42% from 15) of white needles from 1:1 ethyl acetate-petroleum ether (b.p. 30-60°). A sample was recrystallized from methanol-water as feathery needles, m.p. 181.5-183°; the D form⁹ had m.p 180.5-181° and the i.r. spectra were identical.

Anal. Calc. for $C_{14}H_{17}N_5O_3$: C, 55.47; H, 5.65; N, 23.09. Found: C, 55.43; H, 5.86; N, 22.92.

9-(5,6-Dideoxy-α-L-lyxo-hex-5-enofuranosyl)adenine (19). — Compound 18 (500 mg) was treated with 0.05 m sulfuric acid at room temperature for 5 days as described for the D form⁹. The product was separated from unreacted starting-material by chromatography³³ on a column (1.8 × 17 cm) of Bio-Rad AG1-X2 (OH⁻, 200-400 mesh) ion-exchange resin. The column was eluted with 30% aqueous

methanol and 16-ml fractions were collected. Fractions 3 and 4 yielded 14 mg of 18, m.p. 182°. Fractions 23-63 yielded product 19, which was crystallized from water (270 mg), m.p. $247-249^{\circ}$ (dec.), $[\alpha]_{D}^{24} - 52^{\circ}$ (c 1.12, M hydrochloric acid); lit. 7: m.p. 246-247°, $[\alpha]_{D}^{27} - 52.1^{\circ}$ (c 0.305, M hydrochloric acid); the m.m.p. and i.r. spectra confirmed the identity of 19.

Anal. Calc. for $C_{11}H_{13}N_5O_3$: C, 50.18; H, 4.98; N, 26.60. Found: C, 49.91; H, 4.93; N, 26.38.

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REFERENCES

- M. J. ROBINS AND R. A. JONES, J. Org. Chem., 39 (1974) 113-115; M. J. ROBINS AND E. M. TRIP, Tetrahedron Lett., (1974) 3369-3372.
- 2 J. R. McCarthy, Jr., M. J. Robins, L. B. Townsend, and R. K. Robins, J. Amer. Chem. Soc., 88 (1966) 1549-1553; J. P. Horwitz, J. Chua, M. A. Darooge, M. Noel, and I. L. Klundt, J. Org. Chem., 31 (1966) 205-211.
- 3 J. ŽEMLIČKA, R. GASSER, AND J. P. HORWITZ, J. Amer. Chem. Soc., 92 (1970) 4744–4745;
 J. ŽEMLIČKA, J. V. FREISLER, R. GASSER, AND J. P. HORWITZ, J. Org. Chem., 38 (1973) 990–999.
- 4 J. R. McCarthy, Jr., R. K. Robins, and M. J. Robins, J. Amer. Chem. Soc., 90 (1968) 4993-4999.
- 5 J. P. H. VERHEYDEN AND J. G. MOFFATT, J. Org. Chem., 39 (1974) 3573-3579.
- 6 G. KOWOLLIK, K. GAERTNER, G. ETZOLD, AND P. LANGEN, Carbohydr. Res., 12 (1970) 301-311.
- 7 L. M. LERNER, J. Org. Chem., 37 (1972) 477-481.
- 8 N. TANAKA, T. NISHIMURA, H. YAMAGUCHI, AND H. UMEZAWA, J. Antibiot. Ser. A, 14 (1961) 98-102.
- 9 L. M. LERNER, J. Org. Chem., 37 (1972) 470-473.
- 10 L. M. LERNER, J. Org. Chem., 37 (1972) 473-477.
- 11 A. Bloch, unpublished data.
- 12 M. E. EVANS AND F. W. PARRISH, Carbohydr. Res., 28 (1973) 359-364.
- 13 L. M. LERNER, Carbohydr. Res., 36 (1974) 392-397.
- 14 G. Chaves, O. Haines, and A. H. Haines, Carbohydr. Res., 22 (1972) 205-208; J. S. Brimacombe, J. Minshall, and L. C. N. Tucker, ibid., 31 (1973) 146-150.
- 15 P. Jerkeman, Acta Chem. Scand., 17 (1963) 2769-2771.
- 16 L. M. LERNER, J. Org. Chem., 37 (1972) 4386-4391; P. J. BOON, A. W. SCHWARTZ, AND G. J. F. CHITTENDEN, Carbohydr. Res., 30 (1973) 179-182.
- 17 W. Sowa, Can. J. Chem., 49 (1971) 3292-3298.
- 18 W. Sowa, Can. J. Chem., 50 (1972) 1092-1094.
- 19 G. J. G. CHITTENDEN, Carbohydr. Res., 22 (1972) 491-493.
- B. R. BAKER, R. E. SCHAUB, J. P. JOSEPH, AND J. H. WILLIAMS, J. Amer. Chem. Soc., 77 (1955)
 12-15; J. PROKOP AND D. H. MURRAY, J. Pharm. Sci., 54 (1965) 359-365.
- 21 M. J. ROBINS, W. A. BOWLES, AND R. K. ROBINS, J. Amer. Chem. Soc., 86 (1964) 1251-1252.
- 22 J. R. PARIKH, M. E. WOLFF, AND A. BURGER, J. Amer. Chem. Soc., 79 (1957) 2778-2781.
- 23 M. L. WOLFROM, A. B. FOSTER, P. MCWAIN, W. VON BEBENBURG, AND A. THOMPSON, J. Org. Chem., 26 (1961) 3095-3097.
- 24 L. M. LERNER, Carbohydr. Res., 38 (1974) 328-332.
- 25 L. M. LERNER AND P. KOHN, J. Org. Chem., 31 (1966) 339-341.
- 26 J. DAVOLL AND B. A. LOWY, J. Amer. Chem. Soc., 73 (1951) 1650-1655.
- 27 M. J. TAYLOR, B. D. KOHN, W. G. TAYLOR, AND P. KOHN, Carbohydr. Res., 30 (1973) 133-142.
- 28 J. F. HENDERSON, A. R. P. PATTERSON, I. C. CALDWELL, B. PAUL, M. C. CHAN, AND K. F. LAU, Cancer Chemother. Rep., Part 2, 3 (1972) 71-85.

- 29 L. M. LERNER, B. D. KOHN, AND P. KOHN, J. Org. Chem., 33 (1968) 1780-1783.
- 30 M. L. Wolfrom and A. Thompson, Methods Carbohydr. Chem., 2 (1963) 65-67.
- 31 K. IWADARE, Bull. Chem. Soc. Jap., 17 (1942) 372-376.
- 32 K. Freudenberg, A. Wolf, E. Knopf, and S. H. Zaheer, Ber., 61 (1928) 1743-1750.
- 33 C. A. Dekker, J. Amer. Chem. Soc., 87 (1965) 4027-4029.